

## Examination of survival, physiological parameters and immune response in relation to the thermo-resistant heterosis of hybrid oysters derived from *Crassostrea gigas* and *C. angulata*

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High temperature is an important environmental stressor leading to summer mass mortality of oysters. Significant survival heterosis in summer were observed in hybrids between two phylogenetically closely-related oysters, *Crassostrea gigas* and *C. angulata*, but an explicit understanding of heterosis has been lacking here, we investigated the survival performance, oxygen consumption rate and enzyme activity (i.e.,  $\text{ATPase}$ ,  $\text{ATPase}$ , and  $\text{ATPase}$ ) in *C. gigas* GG, *C. angulata* and their hybrids *C. gigas* × *C. angulata* G and *C. angulata* × *C. gigas* G under acute heat stress containing five levels (15, 20, 25, 30, and 35 °C). Compared with parental strains, the hybrid strains exhibited higher cumulative survival rate at all temperatures, indicating heterosis for thermal tolerance. Cox regression analysis showed that hybrid strains challenged with heat shock exhibited lower hazard ratio and longer expected lifetime. Moreover, in hybrid strains were significantly higher  $P < 0.05$  than those in parental strains under 25 and 30 °C Arrhenius break-point temperature and temperature coefficient  $Q_{10}$  revealed that hybrids had broader temperature range of aerobic metabolism and lower sensitivity to rising temperature. In addition, the activities of  $\text{SOD}$  and  $\text{CAT}$  of hybrid strains were significantly higher than the parental strains at 25 and 30 °C-level treatment after 2 h and at 30 °C-level treatment after 4 h, whereas the  $\text{MDA}$  content were reduced, indicating their stronger antioxidant capacity. Accordingly, the enhanced aerobic capacity and antioxidant ability of hybrids under high temperature may contribute to the improvement of thermal tolerance. Our findings can facilitate our understanding of the physiological and immune mechanisms underlying thermo-resistant heterosis and facilitate the breeding of thermo-resistant oyster varieties.

### 1. Introduction

The Pacific oyster *Crassostrea gigas*, originated from northern Asia, is a hardy and euryhaline species (Oudry et al., 2015; Ghaffari et al., 2016) and has become a key mariculture species globally because of its high growth and yield characteristics (Edgecote and Avis, 2017). In recent decades, however, the farmed *C. gigas* has been heavily affected by recurrent summer mortality syndrome (Sang et al., 2015; Uhrmann et al., 2016), which is caused by multiple factors including elevated temperature, pathogens, and physiologic stress associated with reproduction (Égremont et al., 2016; Olomieu et al., 2017). High temperature facilitates the growth of pathogens, inhibits oyster immune system, making them more susceptible to opportunistic pathogens

(Oletchni et al., 2016; Sang et al., 2015) and is therefore considered to be an important incentive for (Sang et al., 2015). At present, several breeding programs for thermal-tolerant oysters have been successfully implemented to address the problem associated with summer mortality (Ehrshberger et al., 2016; Sang et al., 2015; Juárez et al., 2016) and have been proved to be effective to improve the survival rate of the oyster during summer.

The majority of research on crossbreeding in plants and animals has been conducted to combine desirable traits (Alconer and Macay, 2016). In aquaculture, hybridization has been frequently used to increase environmental tolerances when one parental species has a wide range of adaptation or specific tolerance (Elson and Edgecote, 2016; Ahman et al., 2016). For instance, crosses between *Crassostrea* and

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ile tilapias resulted in a red tilapia with strong salinity tolerance (Im et al., 2019). Hybrid *Haliotis rufescens* × *H. discus hannai* exhibited improved thermostability than both purebred populations (Afaruga de la Ru et al., 2019). *C. angulata*, a closely related species of *C. gigas*, naturally inhabits the low latitude regions of southern China, and is characterized by excellent warm-water adaptation (Juvet et al., 2019). Ghaffari et al. (2019), Jiang et al. (2020) and, interestingly, the reciprocal hybrids between *C. gigas* and *C. angulata* exhibited obvious heterosis in survival during the high-temperature months and thermotolerance (Jiang et al., 2020). However, little is known about the physiological and immunological mechanisms of heterosis in thermotolerance of hybrid oysters.

Thermotolerance assessments of oysters are usually based on the survival curves, which ends with the death of the individuals (Howard and Romero, 2019; Shen et al., 2019). The physiological processes, i.e., respiration and excretion of marine ectotherms are influenced by elevated temperature (Magnuson et al., 2019) and is closely related to their thermal resistance (Lar et al., 2019). Elevated or high temperature increase the organism's oxygen consumption rate, which is replenished by increased ventilation (Jansen et al., 2019). Q<sub>10</sub> or Arrhenius' temperature coefficient is frequently used to evaluate the temperature sensitivity of oysters (Saras et al., 2019). However, extremely high temperature induces oxygen restriction, leading to a respiratory peak, over which anaerobic respiration is stimulated, with a rapid decline in oxygen consumption rate (Örtner, 2019). The temperature at respiration peak, Arrhenius break-point temperature (Lar et al., 2019), may represent the thermotolerance threshold of marine organisms (Jansen et al., 2019).

High temperature could induce organisms to produce excessive endogenous reactive oxygen species, resulting in oxidative damage (Ushcha et al., 2019), signal transduction disorder (Ahman and Ahman, 2019) and increased cellular apoptosis (Limén et al., 2019). Malondialdehyde is the part of lipid oxidation byproduct, and its level is frequently used to measure the degree of oxidative stress (Agoudo et al., 2019). To protect cells from detrimental effects, aquatic animals have evolved antioxidant defense systems to neutralize extra and free radicals (Ang et al., 2019). The system consists of non-enzymatic component and antioxidant enzymes, the latter including the enzymes superoxide dismutase and catalase (Liu, 2019; Lein et al., 2019), which could degrade superoxide into oxygen and water (An et al., 2019). Thus, expression of antioxidant enzyme activities may reflect the immune resistance of aquatic organisms under heat stress (Ahman and Ahman, 2019).

To address this issue, we evaluated the survival, physiological parameters and antioxidant enzyme activity, and *in vitro* in *C. gigas*, *C. angulata* and their reciprocal hybrids under acute thermal stress. The integration of phenotypic, physiological and immunological data is aimed to better understand the heterosis in thermotolerance of hybrids.

## 2. Materials and methods

### 2.1. Oysters

Diallel hybridization between *C. gigas* and *C. angulata* was carried out in June. *C. gigas* were sourced from Gongcheng Handong Province, China (108°12'E, 20°15'N), while *C. angulata* were gathered from Hangzhou Huzhou Province, China (120°15'E, 30°15'N). For each species, eggs from females were pooled and divided equally into two batches. Each batch of eggs was fertilized with a mixture of sperm from *C. gigas* or *C. angulata*. Thus, four crosses were established: GG-*C. gigas* × *C. gigas*, G-*C. gigas* × *C. angulata*, G-*C. angulata* × *C. gigas*, and G-*C. angulata* × *C. angulata*. After months, yearling oysters of GG, G, and reciprocal hybrids G and G were obtained from Gongcheng and transferred to Antaiyuan Hatchery, Handong Province for heat shock treatment. For each strain, approximately 100 individuals with similar size were cleaned to

**Table 1**

Summary for the growth characteristics and condition index of four strains

Strains	Shell height (mm)	Shell weight (g)	Shell-free dry weight (g)	Condition index
GG	± ab	± a	± c	± b
G	± a	± a	± a	± a
G	± ab	± a	± b	± a

eliminate attaching organisms and acclimated in 100 L concrete pond with filtered, aerated seawater (salinity 25 psu, temperature 20 °C) for 7 days before the heat shock treatment. Oysters were fed daily with fresh *Phaeodactylum tricornutum* (10 mg/L) and the animals were not fed one day before heat shock was applied.

### 2.2. Survival performance under acute heat stress

Four polyethylene buckets were used for acute heat stress experiment, each divided into equal sections separated by polyethylene nets, forming a total of 16 trial areas. The experimental design was completely random with treatments using a factorial design (temperature × strains and replicates per treatment). In the trial bucket, the temperature was increased from control temperature (20 °C) to designed temperatures (25 °C, 30 °C, and 35 °C) at a rate of 1 °C/h<sup>-1</sup> using a water bath unit with heaters or water chiller. During heat shock period, four strains (oysters per strain) were transferred directly to the same bucket with designed temperatures. Survival data was recorded every 1 h and stopped at 6 h when the cumulative survival rates of GG, G, G and G were calculated. The oyster was regarded as dead when it did not respond to touch with an anatomic needle, and dead individuals are picked up. The seawater was exchanged daily, and the oysters were fed *P. tricornutum* three times a day.

### 2.3. Physiological parameters determination

Eight oysters from each strain (GG, G and G) with similar size were randomly selected from the acclimation environment to determine oxygen consumption rate. In a closed chamber, oysters were fasted for 12 h before measurement. Each chamber contained one oyster, and every experiment consisted of eight replicates. All chambers were filled with air-saturated seawater at the control temperature (20 °C) and designed temperature (25 °C, 30 °C, and 35 °C) and blocked with lithium paraffin to avoid gas exchange with the external environment. The chamber containing empty shells was included to serve as a control to correct for autogenic trends. During heat shock, chambers were immersed in a thermostatic bath to maintain a constant temperature ± 0.1 °C. Oxygen concentrations were determined at the beginning of heat shock and 1 h later. Usually, the soft body of oyster was separated from the shells and dried at 60 °C to measure the shell-free dry weight (mg g<sup>-1</sup> h<sup>-1</sup>) was calculated as the followings (Ghaffari et al., 2019):

$$OR = \frac{(DO_{0h} - DO_{1h})_{Treat} - (DO_{0h} - DO_{1h})_{Blank}}{DW \times t} \times V$$

where  $DO_{0h}$  and  $DO_{1h}$  are the oxygen concentrations at the beginning and 1 h, respectively. "Treat" represents chambers containing oysters. "Blank" represents chamber does not contain oysters.  $g$  is the shell-free dry weight,  $V$  represents the volume of water,  $t$  represents the measuring time.

The condition index (CI), an indicator of the physiological or nutritional condition of oyster, was calculated as the following (Labe and Lbright, 2019):

$$CI = \frac{\text{shell - free dry weight (g)}}{\text{whole wet weight (g) - shell wet weight (g)}} \times 100$$

#### 2.4. Antioxidant enzyme activity determination

The enzyme activity experiment consists of two parts. After 1 h of heat shock treatment, nine oysters each strain were collected from the 0, 1, and 2 °C buckets to examine the effects of different heat shock temperature on the enzyme activities of four strains. Nine oysters per strain from the 0 °C buckets were sampled at 0, 1, 2, 3, 4, 5, and 6 h to evaluate the effects of different heat shock time on the enzyme activities of four strains. Oysters were gill-clipped and the gills were instantly frozen in liquid nitrogen and kept at -80 °C. All enzyme activity measurements were carried out with three replicates, and the gills from three individuals were pooled as one sample for each replicate. The gills were defrosted in -80 °C and -20 °C refrigerators, followed by blotting on Whatman paper and weighing. Samples were homogenized with ice-cold normal saline 1:1 dilution in an ice bath and then centrifuged at 10,000 rpm for 10 min at 4 °C. The supernatant liquid was gathered and the activity of superoxide dismutase (SOD), catalase (CAT), malondial-

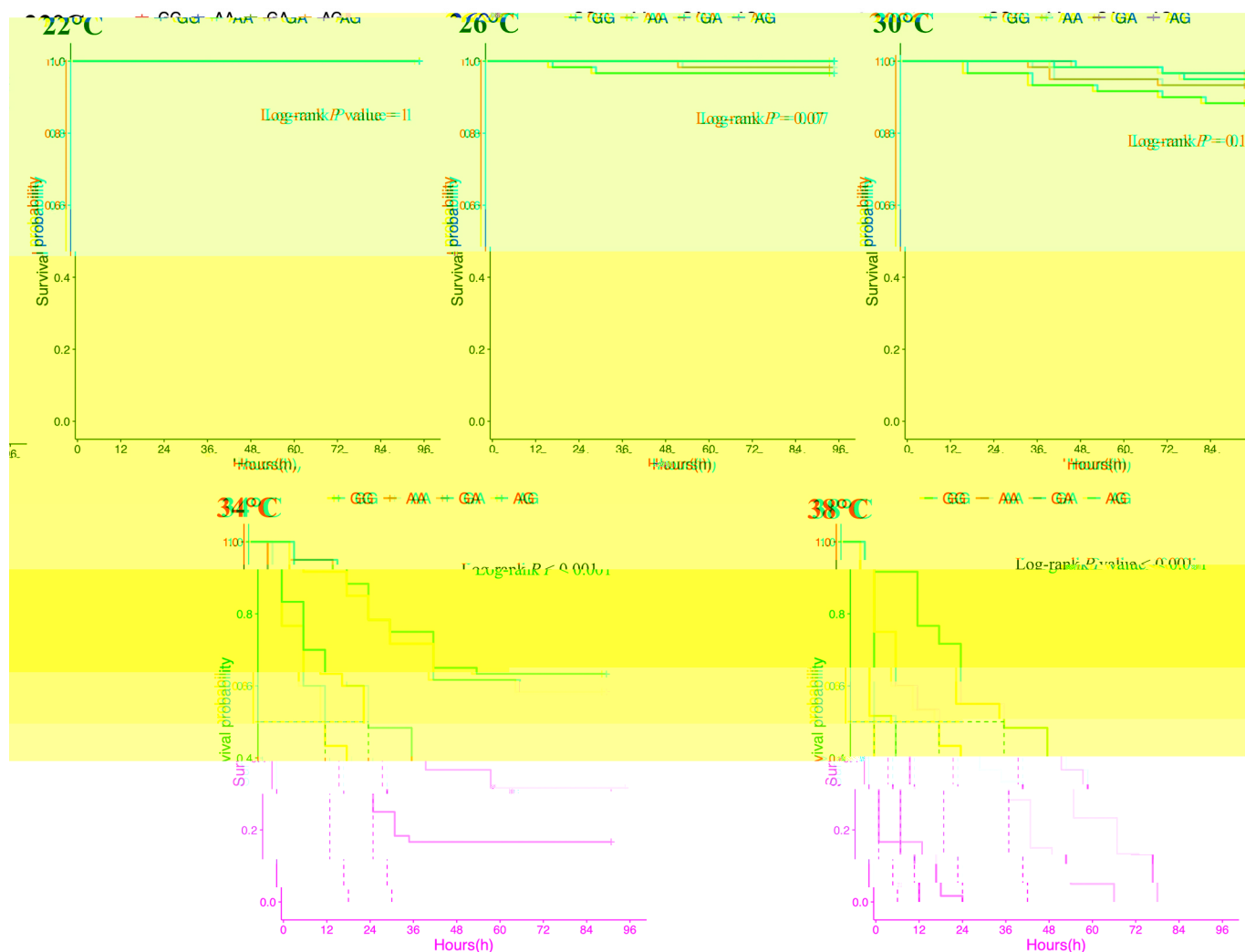


Fig. 1. survival as a function of time of four strains GG, AA, GA and AG under different heat shock temperature levels 22 °C, 26 °C, 30 °C, 34 °C and 38 °C.

Table 2

Box proportional hazard model stratified by heat stress and estimation of expected lifetime (h) for four strains on 22 °C levels

Strains	Coefficient estimate	Hazard ratio	Estimation of expected lifetime
GG	-	-	a
AA	-	-	b
GA	-	-	c
AG	-	-	d

Strains were significantly correlated with survival time ( $P < 0.05$ ). Different lowercase letters in the same line indicating significantly different according to the log-rank test ( $P < 0.05$ ).

Table 3

Two-way analyses of variance testing for temperature by strain interaction effects on oxygen consumption rate and antioxidant enzyme activity (SOD, CAT and GPx).

Effect	Oxygen consumption rate		SOD		CAT		GPx	
	df	F-value	df	F-value	df	F-value	df	F-value
Strain	3	10.21	3	10.21	3	10.21	3	10.21
Temperature	4	10.21	4	10.21	4	10.21	4	10.21
Strain × Temperature	12	10.21	12	10.21	12	10.21	12	10.21
Error	108	-	108	-	108	-	108	-

The P-value associated with each F-value are indicated by asterisks: \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ , - not significant.

strongly affected by strains, temperature and their interaction ( $P < 0.05$ ).  $P < 0.05$ , respectively. In addition, the activity in GG and AA increased gradually as temperature elevated, reaching the maximum 1.2 mgprot<sup>-1</sup> and 1.1 mgprot<sup>-1</sup>, respectively at 30 °C, and declined afterwards. In contrast, the activities in GA and AG continued to rise with increasing temperature, and was significantly higher than the parental strains at 22 °C and 30 °C. Two-way ANOVA revealed clear effects of strain ( $P < 0.05$ ), temperature ( $P < 0.05$ ) and strain × temperature interaction ( $P < 0.05$ ) on the activity levels. A dramatic increase in SOD content for GG (1.2 nmol mgprot<sup>-1</sup>) and AA (1.1 nmol mgprot<sup>-1</sup>) was found over temperature, while no obvious changes in SOD content between both GA and AG up to 30 °C. Interestingly, SOD levels in GA were significantly lower ( $P < 0.05$ ) than the parental

strains at and ° ixed model results indicated signi -  
cant

more stable aerobic metabolism when exposed to high temperatures conditions. Meanwhile, the hybrid strains showed enhanced antioxidant capacity and less oxidative damage at extremely temperature

#### 4.1. Comparison of mortalities under heat stress among cohorts

*C. gigas* from the coast of northern China is recurrently exposed to massive mortality events. It has been estimated that > 50% of *C. gigas* farmed in Shandong were lost in the summer of 2013, when seawater temperatures exceeded 30 °C (Wang et al., 2015). Nonetheless, the survival rate of hybrid oysters was > 90% during warmer months (Jiang et al., 2018). This resistance to high water temperature was supported in the current research, since an increase in temperature from 25 °C to 30 °C leads to an obvious increase in survival rate.

activity or content of the three enzymes  $P < 0.05$  or  $P < 0.01$  (Table 1).

#### 4. Discussion

This study was performed to assess the survival, physiological parameters and immune responses of hybrid oyster under acute heat shock, with an aim of explaining their survival advantage compared to the parental species. We found that the hybrid strains showed larger tolerance to thermal stress since they exhibited higher survival rate and

in a utatic hybrids, such as *Ictalurus punctatus* × *I. furcatus* with low-oxygen tolerance [unham et al ,](#) , *Pomacea canaliculata* × *P. maculate* with low-temperature tolerance [atsu ura et al ,](#) and *Haliotis rubra* × *H. laevigata* with heat tolerance [lter et al ,](#) esides, the ox proportional ha ards analysis revealed that strain, temperature and their interaction had signi cant in uence on cumulative survival rate, indicating the close dependence of strain and temperature on oyster survival his genotype-environment interaction was associated with the biogeographical distribution of parental lines [unham et al ,](#) , which may affect the heat tolerance of hybrids [ahlhoff and omero,](#) s the main *C. gigas* producing area in hina, handong rovince has an average seawater temperature of – ° in contrast, *C. angulata* is a local species in u ian rovince where temperatures are higher, ranging from ° to ° [Jiang et al ,](#) b onse uently, the high heat resistance of hybrid oysters may be inherited from one of its parents, the warm-adapted *C. angulata* [lter et al ,](#) n a similar case in abalone, [iang et al](#) reported *H. discus hannai* × *H. gigantea* successfully inherited the high-temperature resistance of *H. gigantea* owever, in this study, G was more tolerant of high temperature compared to G, which may suggest that the maternal effect was small and the paternal effect was large imilarly, [im ová et al](#) found that the hybrids with common carp in the paternal position exhibited greater viability than the hybrids with common carp in the maternal position

#### 4.2. Change of oxygen consumption in response to heat stress

or several marine animals such as largemouth bass [a et al ,](#) , small clam [Jansen et al ,](#) and esso scallop [Jiang et al ,](#) , ndings showed that metabolic rate was positively associated with seawater temperature owever, a few studies have reported a negative relationship between metabolic rate and environment temperature in some freshwater animals, including limpets [c ahon,](#) and sculpins [alsh et al ,](#) n this study, the of the four oyster strains rst rose and then fell with rising temperature pregu-lated respiration rate was thought to be associated with increased digestion rate and protein synthesis rate [Jiang et al ,](#) , but also indicated the activation energy needed for en yme catalytic reaction was lower at this moment [ochach a and omero,](#) eanwhile, the decrease of may be a protection mechanism against excessive metabolism under high-temperature conditions [Jansen et al ,](#) nimals possessing the capacity to reduce total metabolism was thus thought to be li ely to deal better with extreme temperature than animals without such capacity [iao et al ,](#) n contrast, other studies concluded that species that could maintain their aerobic capacity at higher temperature have a better heat resistance than species that experience a decline in aerobic performance as temperature rises [ing et al ,](#) [Ghaffari et al ,](#) his may be due to the different re- action norms of animals under short-term and long-term stress, allowing them to survive better under different acclimation conditions [Glanville and eebacher,](#) he results in this study were consistent with the latter view, as the hybrids with higher survival rates exhibited higher at – ° urther, G had higher metabolic rate compared with G at ° , indicating stronger aerobic capacity in G under extreme heat n our study, the s were ° and ° for GG and

strain, respectively ne worry is that the measured for GG in this research is close to seawater temperature ° during summer in handong rovince, rendering *C. gigas* vulnerable to temperature rise caused J am□

steadily over temperature or time, which differs from this pattern. Moreover, the increasing activity of with exposure to stress over time was also found in muscle tissue of gold sh *ushcha* and *agnyu ova*, indicating that no switch from aerobic to anaerobic metabolism occurred in the organism (Jiang et al., 2021). In part from that, there were other prominent performances of hybrids may be related to their survival advantage under heat shock. G and G gained significantly higher  $P < 0.05$  activities in and than GG at extremely high temperature and  $0.05 < P < 0.01$ , and activities of G and G were significantly higher than those of GG at  $30 \pm 0.5$  °C. Accordingly, these two characteristics of hybrids may mean earlier response time and higher response intensity of antioxidant system at extreme temperatures, implying an enhanced capacity to maintain cellular functions of the antioxidative system during thermal stress (Ahman and Ahman, 2019; Agoudo et al., 2021). Another striking finding in this research was that the hybrids had lower level of lipid peroxidation and less variation in lipid peroxidation content compared to the parental strains. This is a lipid peroxidation product of polyunsaturated fatty acids in biological membranes and represents the degree of biological oxidative damage. Increased lipid peroxidation content have been reported in whiteleg shrimp (Jiang et al., 2021), common scallop (Jiang et al., 2021) and red swamp crayfish (Guo et al., 2021). In response to short-term heat stress, notably, we found there was a significant negative correlation  $r = -0.85$ ,  $P < 0.001$  between survival and lipid peroxidation in brown scallops, a low-survival population compared to the golden scallops. Thus, the lower lipid peroxidation content of hybrids indicated less oxidative damage, which corresponded with their high survival performance under thermal stress.

## 5. Conclusions

This study demonstrated that the hybrids showed higher cumulative survival rate under thermal stress compared with their parental species, *C. gigas* and *C. angulata*. The significant heterosis of survival rate suggested that crossbreeding improved the thermal tolerance of oysters. Meanwhile, the hybrids exhibited broader temperature range of aerobic metabolism and lower sensitivity to temperature rise, which contribute to the stable and great aerobic capacity of hybrids under high temperature environment. Further, compared with the parental strains, the antioxidant system and activities of hybrids exhibited a more rapid and intensive response. Also, the hybrids maintained lower levels of lipid peroxidation content, indicating less oxidative damage. These findings can facilitate our understanding of the physiological and immune mechanisms underlying heterosis in heat resistance of hybrids and provide a new viewpoint for the future breeding of thermo-resistant oyster.

## CRedit authorship contribution statement

**Gaowei Jiang:** investigation, conceptualization, formal analysis, writing – original draft. **Jianmin Zhou:** investigation. **Geng Cheng:** investigation. **Lingxin Meng:** investigation. **Yong Chi:** investigation. **Chengxun Xu:** supervision. **Qi Li:** supervision, conceptualization, resources, writing – review & editing, funding acquisition.

## Declaration of Competing Interest

The authors declare no conflict of interest.

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